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ROMANIAN KIWIFRUIT BREEDING PROGRAM - PRELIMINARY STUDY OF FIFTEEN MALE HYBRIDS FOR SELECTION AS POLLINATORS

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Abstract

Kiwi is a new fruit that can be grown in Romania. A common Italian-Romanian kiwifruit breeding program was initiated in 1993. For pollinator (male) kiwi plants, breeding programs involve the selection of elite plants with long flowering period and high pollen germination rate. The aim of this study was to evaluate the pollen grains from fifteen Romanian kiwifruit hybrids for identifying the most suitable pollinators for kiwi female selections released from our breeding program. The fresh pollen grains were collected during pollination period (2018-2020), in Petri dishes, maintained at room temperature for 12 hours, and then placed in 15% sucrose solution for germination. At different time interval: 4, 8, 12 and respectively, 24 hours, several measurements were done. Four genotypes – ROP3, ROP6, R2P8 and R3P9, which recorded over 90% germinability after 24 hours, have been selected for further field tests, including artificial pollination and compatibility tests with the kiwi female selections.

Keywords: Actinidia deliciosa, Actinidia chinensis, pollen germination, pollen tube growth, sucrose solution

1. INTRODUCTION

The genus *Actinidia* is native to the humid highlands of south-western China and has spread out to several regions of the world (Ferguson, 2007) regarding his rich nutritional composition of the fruits (Huang et al., 1997). The biochemical properties of the kiwifruit and it's high vitamin C content made it popular among consumers as a healthy fruit (Li et al., 2007). The genus contains more than 50 species and most of them are functionally dioecious, with pistillate and staminate flowers occurring on the separate plants (Ferguson, 1990; Borghezan et al., 2011). Therefore, the fruit set is mostly affected by its own flower biology (Costa, 1993; Snowball, 1996).

Both male and female flowers have both sexes, but staminate vines produce male flowers that contains viable pollen, without functional ovaries and pistillate vines produce female flowers with a functional ovary and non-viable pollen (Schmid, 1978; Howpage et al., 1998; Delaplane and Mayer, 2000). Pistillate flowers are typically larger than staminate flowers, but male vines usually carry more flowers according to Devi et al. (2015).

Pollen grains are small (Goodwin, 1987) and are shed abundantly in clumps from both pistillate and staminate flowers (Schmid, 1978). According to literature, male flowers produce larger quantities of

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pollen grains that insects carry to the stigmas of female flowers (Schmid, 1978; Howpage et al., 1998; Delaplane and Mayer, 2000).

Although flowers are scented, particularly staminate flowers, they are not capable to produce nectar (Schmid, 1978). So, besides insects (especially bees), the wind also contributes to pollination.

Results confirmed that the yield and postharvest quality of fruits are strongly affected by an effective pollination (Stănică and Hoza, 1992; Goodwin et al., 2013). Fertilization of a large proportion of ovules results in bigger kiwifruit (Hopping, 1976). Therefore, fruit size and seed numbers are positively correlated (Pyke and Alspach, 1986; Testolin et al., 1991).

Insufficient pollination leads to unsatisfactory fruit size, shape and uniformity, which, in turn, reduces the market value of the production (Goodwin, 2000). Fruit size is the main factor that determines quality, and together with a proper orchard management, optimal production can only be achieved if conditions for pollination and fruit set are the best (Tromp et al., 2005).

For all these reasons, pollination is a very important and inseparable component in respect of regular and consistent production in a number of fruit crops (Hopping et al., 1982; Costa et al., 1993; Underwood, 2001; Petrisor et al., 2012). The viability, tube growth and morphological homogeneity related to pollen quality are the most important properties in fruit plants (Petrisor et al., 2012). These properties are useful for plant breeders, geneticists, and growers (Bolat and Pirlak, 1999). Relationships between viability and pollen germination have been studied and positive correlation between them was reported by Werner and Chang (1981), Pearson and Harney (1984).

Starting with 1970, according to Davison (1974); Palmer and Clinch (1974), studies of floral biology and pollination of kiwifruit began appearing. Numerous studies on pollen grains have continued and addressed reproductive physiology (Falasca et al., 2010), conservation of germplasm (González-Benito et al., 2004), breeding (Cruz et al., 2008; Chagas et al., 2010; Novo et al., 2010), pollination and fruiting (Nunes et al., 2001; Bettiol Neto et al., 2009). For male kiwi plants in breeding programs, it is important to have ability to produce viable pollen and the flowering periods it is necessary to coincide with the tested fruit producing varieties (Novo et al., 2010).

Several methods can be mentioned for the evaluation of pollen viability: using artificial sucrose media - "Sitting Drop" culture method (Shivanna and Rangaswamy, 1992); evaluation by acetic carmine staining (Domingues et al., 1999); incubation in Baker solution (Oliveira et al., 2001) or the germination test in culture medium (Abreu and Oliveira, 2004; Einhardt et al., 2006; Franzon and Raseira, 2006; Pio et al., 2007).

In Romania, kiwifruit was introduced in 1993, in the South East area through an Italian-Romanian kiwifruit breeding program (Stănică and Zuccherelli, 2007; Stănică, 2009; Stănică and Zuccherelli, 2009). After more than two decades of research, several hybrid genotypes were obtained and introduced to be tested. For pollinator (male) kiwi plants, breeding programs involve the selection of elites with high pollen germination and long flowering period. In previous research (Cotruț et al., 2014) germination rate of few Romanian kiwi genotypes (*Actinidia spp.*) was evaluated after 3, 6 and 9 hours in a culture medium containing 20% sucrose, 5 ppm boric acid (H3BO3) and 1% agar. The results showed that in all kiwi genotypes the germination rate and pollen tube growth varied according to the incubation period and most of the studied genotypes appear to be suitable pollinators.

The aim of this study was to evaluate the pollen grains quality of fifteen kiwifruit hybrids express by: shape index of viable and dead pollen grains, viability percentage (%), germination rate (%) and pollen tubes length (μ m) after 4, 8, 12 and 24 hours. The results were used to identify the most suitable pollinators for kiwi female selections released from our breeding program.

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2. MATERIALS AND METHODS

During 2018-2020 pollination period (early May to beginning of June), flowers from fifteen kiwifruit hybrids (Actinidia spp.) were collected. The biological material studied was represented by 12 male plants and 3 hermaphrodite ones, some of them are presented in Figure 1. The plants were organically grown in Southern Romanian climate, in the Experimental Field of the Faculty of Horticulture, within the University of Agronomic Sciences and Veterinary Medicine of Bucharest, and were trained on a T-bar trellis system.



Figure 1. Male flowers from kiwifruit hybrids (Actinidia spp.)

Anthers from picked flowers were kept in Petri dishes, in the laboratory at room temperature, for 12 hours (Cociu and Oprea, 1989). After the anthers dried and dehisced, the obtained pollen grains were placed in a 15% sucrose solution for germination (Mangalore et al., 2017). The solution was prepared by dissolving 15 grams of sucrose in 100 ml of distilled water (Figure 2).



Figure 2. Anthers from picked flowers kept in Petri dishes and samples preparation steps

After 4, 8,12 and respectively 24 germination hours, several measurements (pollen grains length and width; pollen tube length) and observations (pollen viability; germination rate) were made with Leica DM 1000 LED Microscope equipped with Leica DFC 295 Camera and LAS Core software (Figure 3). Length and width were measured for both viable and dead pollen grains, and expressed in µm. Also pollen tube length was expressed in µm. Germination rate was expressed as a percentage and was calculated by dividing the number of germinated pollen grains per view field by the total number of pollens per view field (Mangalore et al., 2017).

The pollen viability was expressed as a percentage and was determined as the sum of all viable pollen grains per view field divided by the sum of all pollen grains per view field (Mangalore et al.,

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2017). Dead pollen grains were recognized because of the darker colour and smaller dimension than viable pollen grains (Stănică and Hoza, 1992; Mangalore et al., 2017).



Figure 3. Leica DM 1000 LED Microscope equipped with Leica DFC 295 Camera and LAS Core software

The average germination percentage and viability percentage for each kiwifruit hybrid was calculated from five replications (five different overviews from the microscope slide). At least thirty pollen grains were chosen randomly to determine the mean tube length, the mean length and width of viable and dead pollen.

3. RESULTS AND DISCUSSIONS

The plants evolution growth stages developed differently depending on the temperatures and climatic conditions recorded in every year. So, the BBCH 60-69 stages, were registered with a delay of about 1-2 weeks, in 2019 compared to 2018 and 2020. The flowering period ensued between the first decade of May and the first decade of June, for most hybrids (Figure 4). The genotype with the earliest flowering was R0P7, and the longest flowering period was recorded for the hybrid R2P8.

The flowering period of the kiwi plants is quite long, 3-5 weeks, because not all flowers bloom at the same time, especially for varieties with several flowers in inflorescence. It is known that the stigma of female flowers is viable for 7-9 days (Sale, 1981; Goodwin, 2000) from the beginning of flower opening, so a prolonged flowering of male varieties helps to better fertilize the ovaries.

The kiwi fruits size and weight at harvest depend especially on the number of seeds fertilized during pollination period (Stevens and Forsyth, 1982; Anon, 1983; Woolley et al., 1988). Also, the fruits size and weight are affected by management factors such as irrigation, fertilization, the ratio between vegetative growth and the number of fruits, the plantation microclimate etc. (Clinch, 1984; Lees, 1986; Sale, 1986; Lawes et al., 1990; Patterson et al., 1999; Goodwin, 2000; McPherson et al., 2001). The number of seeds is in turn closely related to the number of viable pollen grains deposited on the stigma, so the selection of male plants with a long flowering period and a high quality of germination of pollen grains is a priority among breeding programs (Hopping, 1981; Hopping, 1985; Gonzalez et al., 1994).

Studying the pollen grains shape, in Table 1 can be observed that most of them are ellipsoidal, as Schmid (1978), Dickison et al. (1982), Ferguson and Pusch (1991) mentioned in their research.

According to previous studies, dead pollen grains are smaller that viable ones (Korkutal et al. 2004; Devi et al., 2015) and our results confirmed so (Table 1). In Figure 5 can be observed that dead pollen grains have a darker color compared to viable ones.

Kiwifruit	April			Мау			June		
hybrids			decade						
,	II	III	I	II	III	I	II	III	I
						2			
R0P3 (්)						2			
					•	2			
R0P4 (ợ)									
R0P5 (¢)									
						>			
R0P6 (්)									
					•				
B0P7 (🐴)									
R0P9 (0)									
						K			
R0P14 (♂)									
					•				
R1P5 (්)							\rightarrow		
R1P7 (ợ)							\rightarrow		
R1P10 (්)									
· (0)						•			
B2P2 (2)						· · · · ·			
						· ·			
							\leq		
K2P5 (8)									
						-			
2020 - 4-									
R2P8 (♂)									
R3P2 (්)							>		
R3P9 (♂)							\rightarrow		
							\rightarrow		
						1	▶		
2018	\rightarrow	•	1		1	1	ı		
2019	\rightarrow	•							
2020	\rightarrow	•							

Figure 4. The flowering period of the studied hybrids

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V::f	Viable pol	llen grains	Dead pollen grains			
hybrids	length	width (um)	length (um)	width (um)		
R0P3 (♂)	1.864 ± 0.252	1.573 ± 0.211	1.596 ± 0.112	0.920 ± 0.081		
R0P4 (\$)	2.021 ± 0.296	1.758 ± 0.296	1.927 ± 0.227	1.103 ± 0.094		
R0P5 (\$)	2.172 ± 0.293	1.841 ± 0.327	1.941 ± 0.317	1.339 ± 0.305		
R0P6 (ð)	1.849 ± 0.242	1.740 ± 0.153	1.742 ± 0.286	1.474 ± 0.417		
R0P7 (♂)	1.969 ± 0.147	1.722 ± 0.143	1.704 ± 0.376	1.212 ± 0.042		
R0P9 (♂)	1.741 ± 0.122	1.154 ± 0.259	1.659 ± 0.272	0.930 ± 0.037		
R0P14 (♂)	1.957 ± 0.134	1.356 ± 0.298	1.569 ±0.186	1.084 ±0.124		
R1P5 (♂)	2.017 ± 0.227	1.623 ± 0.173	1.011 ± 0.124	0.780 ± 0.100		
R1P10 (♂)	2.109 ± 0.124	1.756 ±0.056	1.525 ± 0.347	0.912 ± 0.047		
R2P2 (♂)	1.884 ± 0.163	1.508 ± 0.149	1.474 ± 0.232	1.048 ± 0.098		
R2P8 (♂)	1.970 ± 0.171	1.751 ± 0.163	1.893 ± 0.137	1.376 ± 0.122		
R3P2 (♂)	1.874 ± 0.116	1.633 ± 0.157	1.815 ±0.149	1.240 ± 0.028		
R3P9 (♂)	1.774 ± 0.169	1.639 ± 0.219	1.674 ± 0.197	1.294 ± 0.120		

Table 1. Kiwifruit pollen grains measurements (µm)

Note: Mean values ± Standard Deviation



Figure 5. Viable (A) and dead (B) pollen grains of R0P7 male kiwifruit hybrid, after immerging in a 15% sucrose solution

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In Figure 6, can be observed that all studied hybrids recorded over 90% pollen viability, except the hermaphrodite ones R0P4 and R0P5, which recorded 72.48 % and respectively 65.34 %, in 2019, and 64.49 % and respectively 57.47 %, in 2020.



Figure 6. Kiwifruit hybrids pollen viability rate, 2019-2020

According previous studies (Werner and Chang, 1981; Pearson and Harney, 1984), a positive correlation between pollen viability and pollen germination can be observed. For all studied hybrids, pollen germination rates were gradually increasing after 4, 8, 12 and 24 hours. The highest values of the germinability rate, after 24 hours, were recorded at hybrids R2P2 – 90,27%, R0P3 – 90.79 %, R0P6 – 91.17 %, R1P5 – 91,22%, R1P10 – 91,96%, R3P9 – 93.23 % and R2P8 – 93.35 % (Figure 7).



Figure 7. Kiwifruit hybrids pollen germination rate at different time intervals

The results showed that pollen germination rate and pollen tube length were strongly influenced by the genotype and varied also according to the incubation period. Pollen tube lengths for each hybrid increased in time, after 4, 8, 12 and respectively 24 hours (Table 2). After 24 hours of germination, the highest values of pollen tube length were recorded at R3P9 (\circlearrowleft) – 5.440 µm, flowed by R2P8

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 $(\Im) - 4.805 \,\mu\text{m}$, while the lowest value was found at R0P4 (\Im) – 1.460 μ m. The standard deviation showed significant differences between pollen tube lengths in the same view field for all genotypes (Figure 7).

TZ:	Pollen tube length after							
hybrids	4 hours (µm)	8 hours (µm)	12 hours (µm)	24 hours (µm)				
R0P3 (d)	1.713 ± 0.952	2.327 ± 0.455	2.489 ± 0.617	4.206 ± 0.978				
R0P4 (¢)	0.834 ± 0.396	1.197 ± 0.410	1.364 ± 0.440	1.460 ± 0.876				
R0P5 (\$)	1.227 ± 0.776	1.384 ± 0.545	1.455 ± 0.470	2.977 ± 2.056				
R0P6 (3)	0.745 ± 0.328	1.250 ± 0.538	1.397 ± 0.563	1.754 ± 1.623				
R0P7 (ථ)	0.814 ± 0.246	0.818 ± 0.427	1.351 ± 0.346	1.659 ± 0.736				
R0P9 (3)	0.265 ± 0.002	0.903 ± 0.614	1.121 ± 0.233	1.626 ± 0.137				
R0P14 (♂)	0.627 ± 0.316	0.781 ± 0.531	1.411 ± 0.008	1.681 ± 0.085				
R1P5 (ථ)	1.395 ± 0.593	1.808 ± 0.604	1.928 ± 0.823	2.398 ± 0.889				
R1P10 (♂)	1.514 ± 0.093	1.526 ± 0.266	1.656 ± 0.079	1.781 ± 0.479				
R2P2 (♂)	1.388 ± 0.532	1.879 ± 0.886	2.682 ± 0.954	3.396 ± 0.960				
R2P8 (♂)	1.291 ± 0.441	1.388 ± 0.542	2.320 ± 0.701	4.805 ± 0.834				
R3P2 (♂)	0.977 ± 0.540	1.681 ± 0.764	2.343 ± 0.870	4.018 ± 0.794				
R3P9 (♂)	1.593 ± 0.605	1.627 ± 0.690	1.908 ± 0.579	5.440 ± 0.816				

Table 2. Kiwifruit hybrid pollen tube length (mean \pm SD in μ m) during 24 hours

Note: Mean values ± Standard Deviation



Figure 7. Pollen tube length of R2P8 kiwifruit male hybrid after 8 hours of germination

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In the present study we found out that the kiwifruit pollen registered the highest germination rate after 24 hours, using 15% sucrose solution (according also with the previous studies – Mangalore et al., 2017).

4. CONCLUSIONS

Regarding the study of male plants flowering period, it can be mentioned that most kiwi hybrids bloomed between the first decade of May and the first decade of June. The R0P7 hybrid ensued the BBCH 60 stage in the last decade of April, with the earliest development and the longest flowering period was recorded for the R2P8 hybrid.

Results showed that in all kiwi genotypes the germination rate and pollen tube growth varied according to the incubation period. The highest percentage of germination (93%) was recorded after 24 hours of incubation for R2P8 and R3P9. Because the evaluation of pollen germination rate is an essential criterion for kiwi pollinator's characterization, from the eight male hybrids, four genotypes – R0P3, R0P6, R2P8 and R3P9 (which recorded over 90% germinability rate after 24 hours), have been selected for further field tests. Besides artificial pollination and field compatibility tests with the female kiwifruit plants, other ploidy studies have to be done. Most of our kiwifruit hybrid genotypes have been obtained from interspecific crosses (*A. deliciosa* x *A. chinensis*) and probably have different ploidy laves. For a good pollination rate, male and female plants need to have the same ploidy levels. In order to match the right level of ploidy for male-female couples, further studies using the flow cytometry have to be completed.

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6. REFERENCES

- Abreu, I., Oliveira, M. (2004). Fruit production in kiwifruit (Actinidia deliciosa) using preserved pollen. Australian Journal of Agricultural Research, 55, 565–569.
- Anon., 1983. Artificial pollination of kiwifruit. New Zealand Ministry of Agriculture and Fisheries Agricultural Research Division Annual Report 1982/83, 14.
- Bolat, I., Pirlak, L. (1999). An Investigation on Pollen Viability, Germination and Tube Growth in Some Stone Fruits. *Tr. J. of Agriculture and Forestry*, 23, 383–388.
- Borghezan, M., Clauman, A.D., Steinmacher, D.A., Guerra, M.P., Orth, A.I. (2011). In vitro viability and preservation of pollen grain of kiwi (Actinidia chinensis var. deliciosa (A. Chev.) A. Chev). Crop Breeding and Applied Biotechnology (Brazilian Society of Plant Breeding), 11, 338-344.
- Chagas, E.A., Pio, R., Chagas, P.C., Pasqual, M., Bettiol Neto, J.E. (2010). Composição do meio de cultura e condições ambientais para germinação de grãos de pólen de porta-enxertos de pereira. *Ciência Rural*, 40: 261–266.
- Clinch, P.G., 1984. Kiwifruit pollination by honeybees, Taurange observations, 1978-1981. New Zealand Journal of *Experimental Agriculture*, 12, 29-38.
- Cociu, V., Oprea, Ș. (1989). Metode de cercetare in ameliorarea plantelor pomicole. Dacia, România.
- Costa, G., Testolin, R., Vizzotto, G. (1993). Kiwifruit pollination: An unbiased estimate of wind and bee contribution. *New Zealand Journal of Crop and Horticultural Science*, 21(2), 189–195.
- Cotruț, R., Drăghici, E.M., Stănică, F. (2014). Evaluation of pollen germination capacity of some kiwi genotypes (Actinidia spp.). Analele Universității din Craiova Biologie, Horticultura, Tehnologia Prelucrarii Produselor Agricole, Ingineria Mediului, 19, 125–130.
- Cruz, T.V., Souza, M.M., Roza, F.A., Viana, A.J.C., Belo, G.O., Fonseca, J.W.S. (2008). Germinação in vitro de grãos de pólen em *Passiflora suberosa* L. para sua utilização em hibridação interespecífica. *Revista Brasileira de Fruticultura*, 30, 875–879.

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ISSN: 2284-953X
ISSN-L: 2284-9521

Current Trends in Natural Sciences (CD-Rom) ISSN: 2284-9521 ISSN-L: 2284-9521

- Davison, R.M. (1974). Flowering of kiwifruit. Proceedings of kiwifruit seminar, Tauranga. Ministry of Agriculture and Fisheries, New Zealand, 13–16.
- Delaplane, K., Mayer, D. (2000). Crop pollination by bees. New York: CABI Publishing, 344.
- Devi, I., Thakur, B.S., Garg, S. (2015). Floral morphology, pollen viability and pollinizer efficacy of kiwifruit. International Journal of Current Research and Academic Review, 3 (8), 188-195.
- Dickison, W.C., Nowicke J.W., Skvarla J.J. (1982). Pollen morphology of the *Dilleniaceae* and *Actinidiaceae*. *American Journal of Botany*, 69, 1055-1073.
- Domingues, E.T., Tulmann Neto, A., Teófilo Sobrinho, J. (1999). Viabilidade do pólen em variedades de laranja doce. *Scientia Agrícola*, 56, 265–272.
- Einhardt, P.M., Correa, E.R., Raseira, M.C.B. (2006). Comparação entre métodos para testar a viabilidade de pólen de pessegueiro. *Revista Brasileira de Fruticultura*, 28, 5–7.
- Falasca, G., Franceschetti, M., Bagni, N., Altamura, M.M., Biasi, R. (2010). Polyamine biosynthesis and control of the development of functional pollen in kiwifruit. *Plant Physiology and Biochemistry*, 48, 565–573.
- Franzon, R.C., Raseira, M.C.B. (2006). Germinação in vitro e armazenamento do pólen de Eugenia involucrata DC (Myrtaceae). *Revista Brasileira de Fruticultura*, 28, 18-20.
- Ferguson, A.R. (1990). The genus Actinidia. In Warrington IJ, Weston GC (Eds.), Kiwifruit: Science and management. Auckland, New Zealand: The New Zealand Society of Horticultural Science.
- Ferguson, A.M., Pusch, W.M. (1991). Development of mechanical dry-pollen application to kiwifruit. *ISHS, Kiwifruit II, Acta Horticulturae*, 297, 299-304.
- Ferguson, A.R. (2007). The need for characterization and evaluation of germplasm: kiwifruit as an example. *Euphytica*, 154, 371382.
- Gonzalez, M.V., Coque, M., Herrero, M. (1994). Pollinator selection in kiwifruit (Actinidia deliciosa). Journal of Horticultural Science, 69, 697-702.
- Gonzalez-Benito, M.E., Clavero-Ramírez, I., López-Aranda, J.M. (2004). The use of cryopreservation for germplasm conservation of vegetatively propagated crops. *Spanish Journal of Agricultural Research*, 2, 341-351.
- Goodwin, R.M. (1987). Ecology of honey bee (Apis mellifera L.) pollination of kiwifruit. Thesis, University of Auckland, New Zealand.
- Goodwin, R.M. (2000). Zespri Innovation Kiwifruit Pollination Manual. Zespri Innovation Company Limited & The Horticulture and Food Research Institute of New Zealand Limited.
- Goodwin, R.M., McBrydie, H.M., Taylor, M.A. (2013). Wind and honey bee pollination of kiwifruit (Actinidia chinensis 'HORT16A'). New Zealand Journal of Botany, 51(3), 229-240.
- Huang, H., Dane, F., Wang, Z.R., Jiang, Z.W., Huang, R.H., Wang, S. (1997). Isozyme inheritance and variation in *Actinidia. Heredity*, 78, 328-336.
- Hopping, M.E. (1976). Effect of exogenous auxins, gibberellins and cytokinins on fruit development in Chinese gooseberry (*Actinidia chinensis* Plunch.). New Zealand Journal of Botany, 14, 69–75.
- Hopping, M., 1981. Kiwifruit: hand pollination to improve fruit size. The Orchardist of New Zealand, 54: 258.
- Hopping, M.E., Simpson, L.M. (1982). Supplementary pollination of tree fruit. *New Zealand Journal of Agricultural Research*, 25, 245-50.
- Hopping, M.E. (1985). Kiwifruit pollination. New Zealand MAF Agricultural Research Division Annual Report 1984/85, New Zealand Ministry of Agriculture and Fisheries, 63.
- Howpage, D., Vithanage, V., Spooner-Hart, R. (1998). Pollen tube distribution in the kiwifruit (Actinidia deliciosa) pistil in relation to its reproductive process. *Ann. Bot.*, 81, 697-703.
- Korkutal, I., Kok, D., Bahar, E., Sarkaya, C. (2004). Determination of flower morphologies and phenologies in Hayward and Matua kiwifruit (*Actinidia deliciosa*) cultivars. *Ziraat Fakultesi Dergisi Akdeniz Universitesi*, 17, 217-224.
- Lawes, G.S., Woolley, D.J., Lai, R. (1990). Seeds and other factors affecting fruit size in kiwifruit. Acta Horticulturae, 282, 257-264.
- Lees, N. (1986). Growing export kiwifruit. Kiwifruit Pollination Seminar Proceedings, Ministry of Agriculture and Fisheries, Tauranga, New Zealand, 6.
- Li, Z., Kang, M., Huang, H., Testolin, R., Jiang, Z., Li, J., Wang, Y., Cipriani, G. (2007). Phylogenetic relationships in *Actinidia* as revealed by nuclear DNA genetic markers and cytoplasm DNA sequence analysis. *Acta Horticulturae*, 753, 45–58.
- Mangalore, C., Negi, M., Pant, S.C. (2017). Pollen studies in kiwifruit (Actinidia deliciosa Chive.). International Journal of Agricultural Science and Research, 7 (2), 15-22.

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- McPherson, H.G., Richardson, A.C., Snelgar, W.P., Patterson, K.J., Currie, M.B. (2001). Flower quality and fruit size in kiwifruit (*Actinidia deliciosa*). New Zealand Journal of Crop and Horticultural Science, 29(2), 93-101.
- Novo, M., Romo, S., Rey, M., Prado, M.J. and González, M.V. (2010). Identification and sequence characterization of molecular markers polymorphic between male kiwifruit (*Actinidia chinensis* var. *deliciosa* (A. Chev.) A. Chev.) accessions exhibiting different flowering time. *Euphytica*, 175, 109-121.
- Oliveira, M.S.P., Maués, M.M., Kalume, M.A.A. (2001). Viabilidade de pólen in vivo e in vitro em genótipos de açaizeiro. *Acta Botânica Brasilica*, 15, 27-33.
- Palmer-Jones, T., Clinch, P.G. (1974). Observations on the pollination of Chinese gooseberries variety 'Hayward'. *New Zealand journal of experimental agriculture*, 2, 455-458.
- Patterson, K.J., Snelgar, W.P., Richardson, A.C., McPherson, H.G. (1999). Flower quality and fruit size of Hayward kiwifruit. *Acta Horticulturae*, 498, 143-150.
- Pearson, H.M., Harney, P.M. (1984). Pollen viability in Rosa. HortScience, 19(5), 710-711.
- Petrisor, C., Mitre, V., Mitre, I., Jantschi, L., Balan, M.C. (2012). The Rate of Pollen Germination and the Pollen Viability at Ten Apple Cultivars in the Climatic Conditions of Transylvania. *Bulletin UASVM Horticulture*, 69(1), 417-418.
- Pio, L.A.S., Ramos, J.D., Pasqual, M., Junqueira, K.P., Santos, F.C., Rufini, J.C.M. (2007). Viabilidade do pólen de laranjas doces em diferentes condições de armazenamento. *Ciência e Agrotecnologia*, 31, 147-153.
- Pyke, N.B., Alspach, P.A. (1986). Inter-relationships of fruit weight, seed number and seed weight in kiwifruit. *New Zealand Journal of Agricultural Science*, 20, 153–156.
- Sale, P.R. (1981). Kiwifruit Pollination, male to female ratios and bee activity. Aglink HPP233, New Zealand Ministry of Agriculture and Fisheries, 3.
- Sale, P.R. (1986). Factors which influence fruit size. Kiwifruit Pollination Seminar Proceedings, Ministry of Agriculture and Fisheries, Tauranga, New Zealand, 6.
- Schmid, R. (1978). Reproductive anatomy of Actinidia chinensis (Actinidiaceae). Botanischer Jahrbucher fur Systematick, Pjlanzengeschichte und Pjlanzengeographie, 100(2), 149-195.
- Shivanna, K.R., Rangaswamy, N.S. (1992). Pollen-Pistil Interaction. In: Pollen Biology, Springer. Berlin: Heidelberg.

Snowball, A. (1996). The timing of flower evocation in kiwifruit. J. Hort. Sci., 71, 335-347.

- Stănică, F., Hoza, D. (1992). The pollen's viability at some varieties of chinese japanese plumtree (Germinabilitatea polenului la unele soiuri de prun chino-japonez). Bioterra 1, Bulletin of Scientific Information, Athenaeum Academic University, Faculty of Horticultural Sciences and Bioengineering, 49-52.
- Stănică, F., Zuccherelli, G. (2007). New selections of *Actinidia arguta* from the Romanian breeding program. *Acta Horticulturae*, 753: 263–267. Doi: 10.17660/ActaHortic.2007.753.32.43.
- Stănică, F. (2009). Kiwifruit, the fruit of XXth Century. Lucrări științifice USAMVB, Seria B, 53, 15–28.
- Stănică, F., Zuccherelli, G. (2009). Nuove selezioni di Actinidia arguta dal programma di miglioramento genetico italoromeno. Societa Orticola Italiana, Italus Hortus Journal, 16, 262-265.
- Stevens, I., Forsyth, M. (1982). Pollination, fertilisation, and fruit development. *Proceedings of Kiwifruit Pollination* Seminars, Ministry of Agriculture and Fisheries, Hamilton & Tauranga, New Zealand, 2-10.
- Testolin, R., Vizzotto, G., Costa, G. (1991). Kiwifruit pollination by wind and insects in Italy. *New Zealand Journal of Crop and Horticultural Science*, 19, 381–384.
- Tromp, J., Webster, A., Wertheim, S. (2005). Fundamentals of temperate zone tree fruit production. *Leiden: Backhuys Publishers*, 345.
- Werner, D.J., Chang, S. (1981). Stain testing viability in stored peach pollen. HortScience, 16(4), 522-523.
- Woolley, D.J., Lawes, G.J., Lai, R. (1988). Factors affecting fruit size. New Zealand Kiwifruit Authority National Research Conference, New Zealand Kiwifruit Special Publication, Rotorua, 2, 11-14.
- Underwood, R. (2001). Pollination of kiwifruit. New Zealand Kiwifruit Journal, 44-46.